

PHLEBOTRICHIN, A PHENOLIC COMPOUND FROM THE FRESH LEAVES OF *VIBURNUM PHLEBOTRICHUM*

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Key Word Index—*Viburnum phlebotrichum*, Caprifoliaceae, phenolic glucoside, arbutin, phlebotrichin, 2,6-dimethyl-6(S)-hydroxyl-2-trans-2,7-octadienoic acid, linalool-1-ol acid

Abstract—Arbutin and a new phenolic glucoside were isolated from the fresh leaves of *Viburnum phlebotrichum*. The structure of the new compound named phlebotrichin was established as 1-O-(4-hydroxyphenyl)-6-O-[(2E, 6S)-2,6-dimethyl-6-hydroxy-2,7-octadienoyl]-β-D-glucopyranoside.

INTRODUCTION

We were interested in the well-known fact that the leaves of plants belonging to the genus *Viburnum* are liable to go brown when they are heated or pressed between papers for preparing dried specimens. This report deals with the isolation of arbutin (1), the isolation and structural determination of a new phenolic compound, phlebotrichin (2), and the quantitative comparison of the phenolic compounds in the fresh and brown leaves of *Viburnum phlebotrichum* Sieb. et Zucc.

RESULTS AND DISCUSSION

The fresh leaves were treated as described in the Experimental to yield 1 and 2. Compound 1, arbutin, was identified by direct comparison with an authentic sample. Compound 2, phlebotrichin, white powder-like crystals, mp 141–143°, C₂₂H₃₀O₉, [α]_D²⁷ = –32.6°. The IR spectrum showed the presence of carbonyl and hydroxyl groups whilst the ¹H NMR spectrum showed the presence of two methyl groups, one vinyl group, A₂B₂-type aromatic protons and an aromatic hydroxyl group. Hydrolysis of 2 with 0.5 N sodium hydroxide gave 3 and 4. Compound 3, a pale-yellow oil, C₁₀H₁₆O₃, was soluble in 5% aq. sodium bicarbonate. Its structure was established as (E)-linalool-1-ol acid (2,6-dimethyl-6-hydroxy-2-trans-2,7-octadienoic acid) by comparison of its IR, NMR and mass spectra with those recently described for the natural [1] and synthetic [2] forms of (E)-linalool-1-ol acid. Compound 4, colourless needles, mp 199–200°, C₁₂H₁₆O₇, was identified as arbutin by IR and mixed fusion.

The ¹H NMR data of 2 were compared with those of 1 and 3 (Table 1). The signals of C-5 and C-6 in 1 were shifted +3.1 and –2.6 ppm respectively compared to those of 2 and the signal of δ 168.1 in 2 indicates that C-1" in 3 was attached to C-6 in 1 by an ester linkage. The signs of specific rotation of 3 ([α]_D²⁵ +19.3°) and molecular rotations of 1 (–174.3°), 2 (–142.9°) and d-linalool (+29.8°) suggested that 2 and 3 had a S-configuration at C-6" (Fig. 1).

From the available data, the structure of 2 was established to be 1-O-(4-hydroxyphenyl)-6-O-[(2E,

Table 1 ¹³C NMR data for compounds 1–3 (C₅H₅N)

Carbon No	2	1	3
1	103.6	103.6	—
2	74.9	75.1	—
3	78.5	78.8	—
4	71.7	71.3	—
5	75.5	78.6	—
6	65.0	62.4	—
1'	154.1	154.0	—
2'	116.8	116.9	—
3'	118.9	118.8	—
4'	151.7	151.8	—
5'	118.9	118.8	—
6'	116.8	116.9	—
1"	168.1	—	170.5
2"	127.7	—	128.7
3"	143.6	—	142.4
4"	24.1	—	24.1
5"	41.5	—	41.7
6"	72.2	—	72.1
7"	146.5	—	146.4
8"	111.8	—	111.6
9"	12.5	—	12.7
10"	28.7	—	28.4

6S)-2,6-dimethyl-6-hydroxy-2,7-octadienoyl]-β-D-glucopyranoside. The contents of 1 and 2 in methanol extracts of fresh and brown leaves were assayed by using a dual wavelength TLC scanner. The results showed that the brown leaves contained only ca 0.04% of 1 and 0.008% of 2, while the fresh leaves contained ca 0.11% and 0.08%, respectively. They suggest that the change in colour of the leaves from green to brown is related to the breakdown of 1 and 2.

EXPERIMENTAL

Mps were uncorr. ¹H NMR 100 MHz, TMS as int. standard. Plant material was collected at Ohme in Tokyo, 1979.

Extraction and isolation Fresh leaves (190g) were extracted

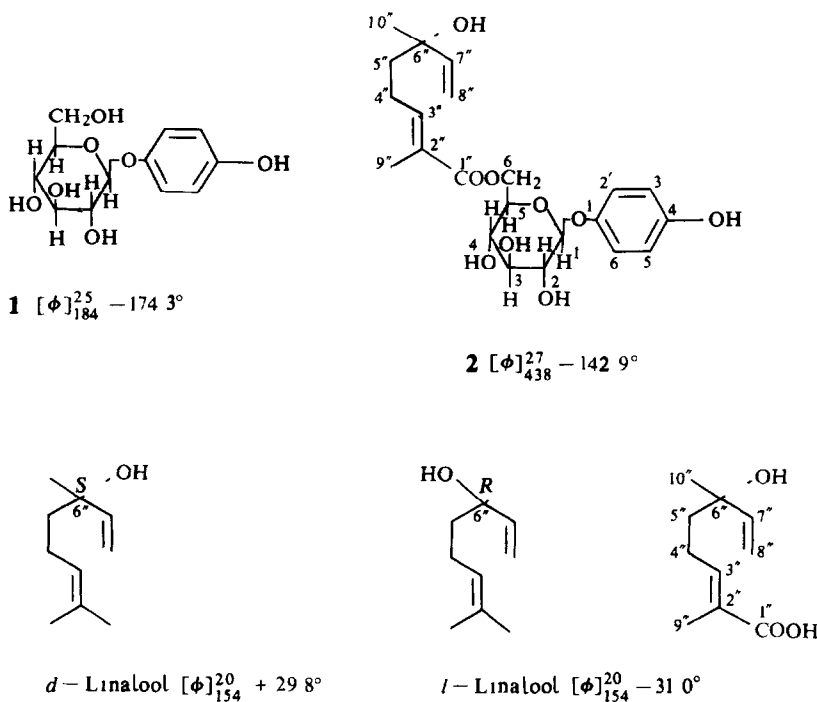


Fig 1 Structures and molecular rotations of 1, 2 and D- and L-linalool

with MeOH (601). The extracts were concd *in vacuo* and diluted with H₂O. The soln was extracted with EtOAc and then *n*-BuOH.

Purification of arbutin (1) and phlebotrichin (2). The *n*-BuOH extract (4.5 g) was subjected to Rotation Locular Counter Current Chromatography (RLCCC) [3] [EtOAc–EtOH–H₂O (10:3:6), upper layer (mobile phase, 242 ml), lower layer (stationary phase, 373 ml), locular 800, flow rate 0.32 ml/min, 1 fraction (fr), 50 ml]. Frs 16–38 were collected, taken to dryness and chromatographed over Si gel using EtOAc–MeOH–H₂O (21:4:3) to give arbutin (415 mg). The EtOAc extract (4 g) was chromatographed on RLCCC [EtOAc–hexane–EtOH–H₂O (10:5:6:5), upper layer (247 ml), lower layer (390 ml), locular 800, flow rate 0.32 ml/min, 1 fr, 50 ml]. Frs 25–40 were collected, taken to dryness and chromatographed over Si gel using EtOAc to afford phlebotrichin (400 mg).

Arbutin (1) was recrystallized (EtOAc) to give colourless needles, mp 199–200°. High resolution MS (70 eV) Found 272.0849, calc for C₁₂H₁₆O₇, 272.2542, UV $\lambda_{\max}^{\text{EtOH}}$ nm (log ϵ) 285 (3.42), IR ν_{\max}^{KBr} cm^{−1} 3300, 1515, 1210, 830, ¹H NMR (Me₂CO-*d*₆) δ 6.73 (d), 6.95 (d), 8.02 (s).

Phlebotrichin (2) was recrystallized (*n*-hexane–EtOAc) to give white powder-like crystals, mp 141–143°, $[\alpha]_D^{27} - 32.6^\circ$ (EtOH, c5). High resolution MS (70 eV) Found 438.1861, calc for

C₂₂H₃₀O₉, 438.4736, UV $\lambda_{\max}^{\text{EtOH}}$ nm (log ϵ) 285 (2.7), IR ν_{\max}^{KBr} cm^{−1} 3400, 1695, 1640, 1510, 1215, 815, ¹H NMR (Me₂CO-*d*₆) δ 1.28 (3H, s, H-10''), 1.83 (3H, s, H-9''), 5.01 (1H, dd, *J* = 2, 10 Hz, H-8''), 5.23 (1H, dd, *J* = 2, 17 Hz, H-8''), 5.98 (1H, dd, *J* = 10, 17 Hz, H-7''), 6.73 (2H, d, *J* = 9 Hz, H-2' or H-6', H-3' or H-5'), 6.95 (2H, d, *J* = 9 Hz, H-5' or H-3', H-6' or H-2'), 8.02 (1H, s, OH-4'), ¹³C NMR See Table 1.

Hydrolysis of phlebotrichin by alkali. A soln of 2 (50 mg) in 0.5 N NaOH (5 ml) was stirred for 10 min at room temp. The soln was acidified (dil HCl) and extracted with Et₂O. The extract was evaporated and chromatographed on Si gel using C₆H₆–EtOAc (5:1) to afford 3 (20 mg). The acidic soln was neutralized (dil aq NaOH) and extracted with EtOAc. The extract was evaporated *in vacuo* to give 4 (25 mg).

Compound 3 was obtained as a pale yellow oil $[\alpha]_D^{25} + 19.3^\circ$ (CHCl₃, c0.15). High resolution MS (70 eV) Found 184.1126, calc for C₁₀H₁₆O₃, 184.2346, IR $\nu_{\max}^{\text{capillary}}$ cm^{−1} 3400, 1680, 1640, 995, ¹H NMR (CDCl₃) δ 1.32 (3H, s, H-10'), 1.64 (2H, dd, *J* = 7, 17 Hz, H-5), 1.82 (3H, s, H-9'), 2.25 (2H, dd, *J* = 7, 17 Hz, H-4'), 5.08 (1H, dd, *J* = 2, 10 Hz, H-8''), 5.24 (1H, dd, *J* = 2, 17 Hz, H-8''), 5.92 (1H, dd, *J* = 10, 17 Hz, H-7''), 6.89 (1H, t, H-3'), ¹³C NMR See Table 1. These spectral properties were identical to those of synthetic (*E*)-linalool-1-oic acid.

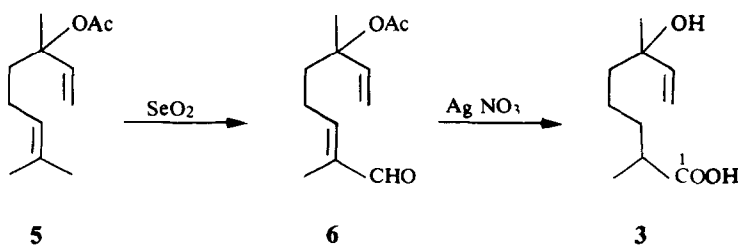


Fig 2 Chemical synthesis of linalool-1-oic acid

Compound **4** was recrystallized from EtOAc to give colourless needles, and identified with arbutin by IR and mixed fusion

Synthesis of (E)-linalool-1-ol acid (3) Compound **3** was synthesized from linalool acetate (**5**) (20 g) by a modification of the method (Fig 2) used for the synthesis of D-2,6-dimethyl-8,8-ethylendioxyoct-2-enioic acid from the cyclic acetal of D-citronellal [2] The aldehyde (**6**) (15 g) was obtained as a colourless oil (bp 124–125°/5 mm) Compound **3** (1.5 g), also obtained as an oil, was purified by chromatography on Si gel in C₆H₆–EtOAc (7/3) **3** pale yellow oil, C₁₀H₁₆O₃ (MS), IR $\nu_{\text{max}}^{\text{capillary}}$ cm⁻¹ 3400 (OH), 1680 (conj COOH), 1640, 1410, 1365, 1280, 1150, 1100, 1065, 995, 920 (CH = CH₂), 800, 740, 685, 640, 550

Quantitative analysis of arbutin and phlebotrichin in fresh and brown leaves Fresh leaves (90 g) and brown leaves which had been prepared from fresh leaves (90 g) by treatment for 3 hr at 30° in a drying apparatus, were extracted with MeOH (6 l) and the extracts reduced (*in vacuo*) in vol to 250 ml Each extract (1 μ l) was then assayed for **1** and **2** using a dual wavelength TLC scanner (Shimadzu CS-910) (TLC, Si gel 60F 254,

EtOAc–MeOH–H₂O (21/4/3), *R_f* arbutin 0.62, phlebotrichin 0.86, 22°, wavelength λ_1 = 290 nm, λ_2 = 340 nm) Fresh leaves arbutin 48.4 mg (0.11%), phlebotrichin 37.9 mg (0.08%), brown leaves arbutin 18.5 mg (0.08%), phlebotrichin 3.6 mg (0.008%)

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